"Welcome to Mars" LB144-Pandemic edition



(Preparing for) **Tuesday's lecture:**

Budgeting homework time (45 min): Ch. 18, section 18.1 (the second half on frogs) is 2166 words in length. At what's considered slow reading speed, 200 words per minute, reading the second half of section 18.1 should take 11 minutes. But when done properly, when you pause to review figures, read and think about a few of the Integrating Questions, and take careful notes, if you focus (avoid distraction) it should take you approx. 45 minutes.

- in your lecture notebook (handwritten notes lead to far greater learning).
- 2. them. Also try to answer the green Review questions.
- 3. microphone so everyone can hear you in lecture) so prepare well.
- 4.

For the second lecture, slowly read the second half of section 18.1 "Frog choruses attract predators." As you read it on your computer or tablet, please be sure to take handwritten notes on paper

_____ Try to answer some Integrating Question and Review Questions. As you read the ICB textbook always attempt to answer at least one of the yellow Integrating Questions each time you get to a set of

(Trifecta): Prepare to explain (aloud) Figures 18.6 and Table 18.2 in class. As you read a section from the ICB textbook always attempt to pause and study each figure/drawing/table that is discussed. In class, during lecture, you may be randomly chosen to explain these aloud (the LA will hand you a

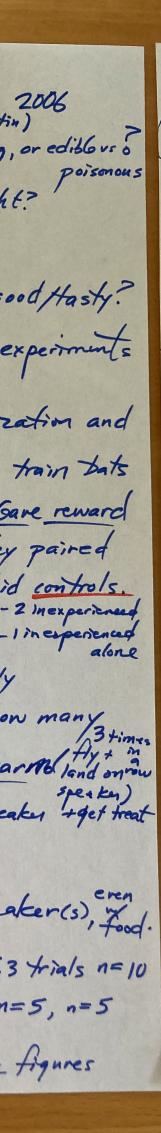
Advanced TIP: scroll down to the bottom of the page, in the Bibliography, and click on the link to an original paper by Dr. Rachel Ryan to see which data was used to make figure 18.6 and Table 18.2, and look at Ulagaraj's research paper on crickets to get used to, and in a habit of, checking original papers.



Keading Chapter 18 - section 1 Exploiting communication Detween others -? some species intercept information transfer between members at another L.O.s - identity commonalities tetween communication within or -evaluate how into is used to find/exploit otherspecies species - provide examples Intro - One risk in communitating ... is other species could eat you. parasited - One risk in communitating ... is other species could eat you. parasited prey hosts Example: "Frog choruses attract predators (ornell Milwankee Musum Mike Ryan - studied frogs in Panama. Male frogs vocalize to attract mates, and in large groups. Risk: discovered by enemy. Do predators to frog (opossum + bats) hear or see or what? Study # Ryan et al Researchers did purely observational experiment, watched frogs vs opossums. Saw via night vision scope. Saw 31 emits. Purpose: how opossums hunt? Methods: Observe, scope, record. Findings: n=39 captures, general trend opossum approached edge of pond, turned toward rocalizing frogs, as walked closer stopped agam, turned head side to side rotating cas. Then finally ran forward + pounced attempting a capture.

Did a "playback" experiment. speaker away from pondedy

(Ryan et al, study #2 cont.) Rachel Page + Mike Ryan - bat learning - Univ Texas 2006 (Austin) Q: How doe bats learn which vocalizations = small vs big, or ediblous of poismous M. Playback used speaker, placed 2 meters away from Ponch cage (Nhy?). "Location frogs not even found?" Play recordings of tungara frog vocalizations ONLY when # Know fringe-lipped bat lives in social groups, get taught? a <u>Opposum</u> was near speaken BUT facing away. + not when other frogs were vocalizing. Study # 1 + 2 + 3 Purpose : How /when do bats learn which frug calls one good Hasty? F.) (findings) In 5 out of 5 trials (n=5) oppossum turned Methods: Use came toad vocalization in playback experiments toward speaken, did head tilting / turns + car rotating. Then Cone toad is both too BIG + poisonous to bats. approach speaken + pounced. 3 of 5 times (n=3/2). Used caged pats, in captivity, First just play vocalization and Researchers claim in 2 times not pouncing animal spotted observe if any are attracted (rope). Next attempt to train bats IQ10 think it uses a constic or visual cues to locate prey? Can philonder opossum intercept frog communications? (nocturnal) to become attracted to care toad's call speaker). Gave reward for landing Fon speaker (raw fish treat). Last they paired a bet successfully trained with naire partner + did controls. (experienced) (in experienced) -2 mexperienced -1 in experienced Ryan et al, study #3) Redator #2 fringe-lipped bat vs frogs Purpose: Can predator distinguish between species of frog? Via votatizahn (this bat uses hearing to find poey) Methods (M): They In each frial 1-5 speakers set-up, one rondomly chosen to play cane toad calls. Keep track how many times needed for each mexperienced bat to learne land on ou Visit frog preeding Ponds Jan. to June. Observational Study. Half ponds had Tungara frogs Hasty). Categorized data into amount af group rocalizations: "full chorus" "partial chorus" "few" "none" calling. Recorded # pat "risits" Findings: First, bats never flew to any silent speaker(s), Food. Ultimately, bats learn fast from peers. Figure 18.6 A bats Mill Figure 18.6 A bats Mill Ryan et al Study #4, Play back to captime twild bats vocation for some of edible vs poisonous frog, or small vs oversized. 1818, bab L. I. I.G. II, 12, 13 Table 18.2 mixed pair learned quickley - average 5.3 trials n=10 but controls took long time 96 trials. n=5, n=5 IQ 14,15 Review Bibliography (click + look) see figures



Section 18.1 What is a learning goal from last class (what part of biology are we exploring)?

Biology Learning Objectives

- and exploit other species.
- species.

Section 18.1 Have organisms evolved to exploit communication between individuals of other species?

• Identify the commonalities between communication within a species and communication between species.

• Evaluate how information is used by organisms to find

• Provide examples of adaptations of one species to the information passed between individuals of another

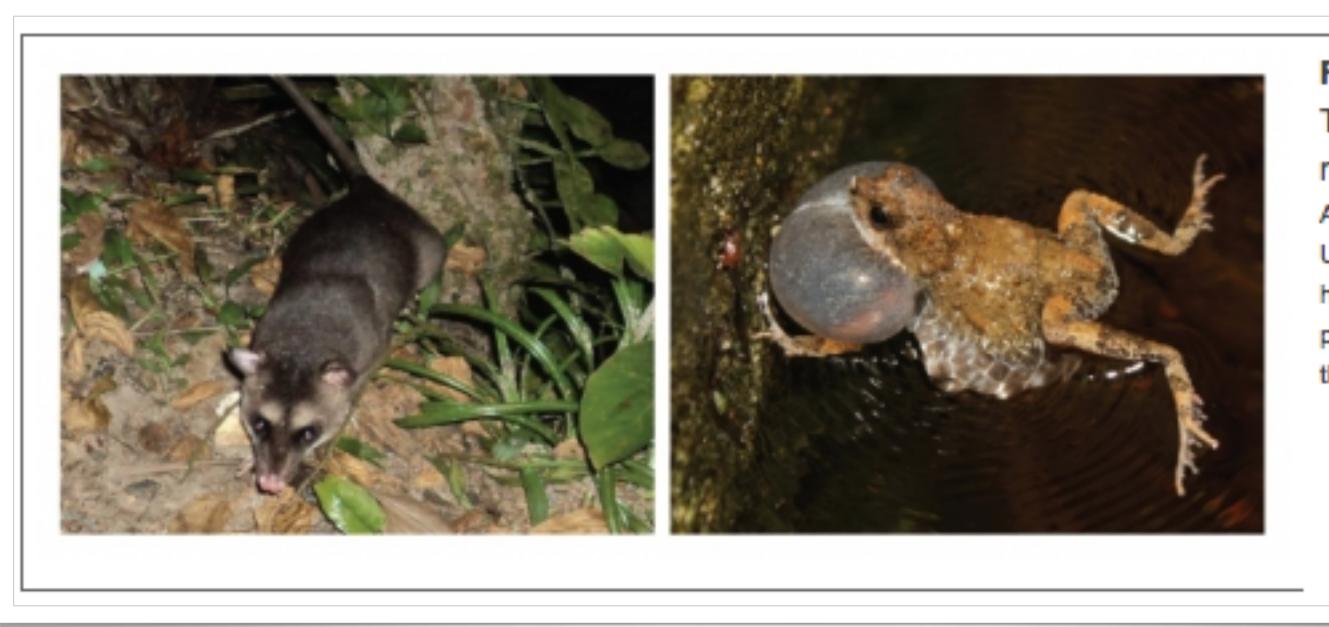
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Section 18.1: What did you find the most interesting from today's reading (how might these readings help you)?

Test your knowledge

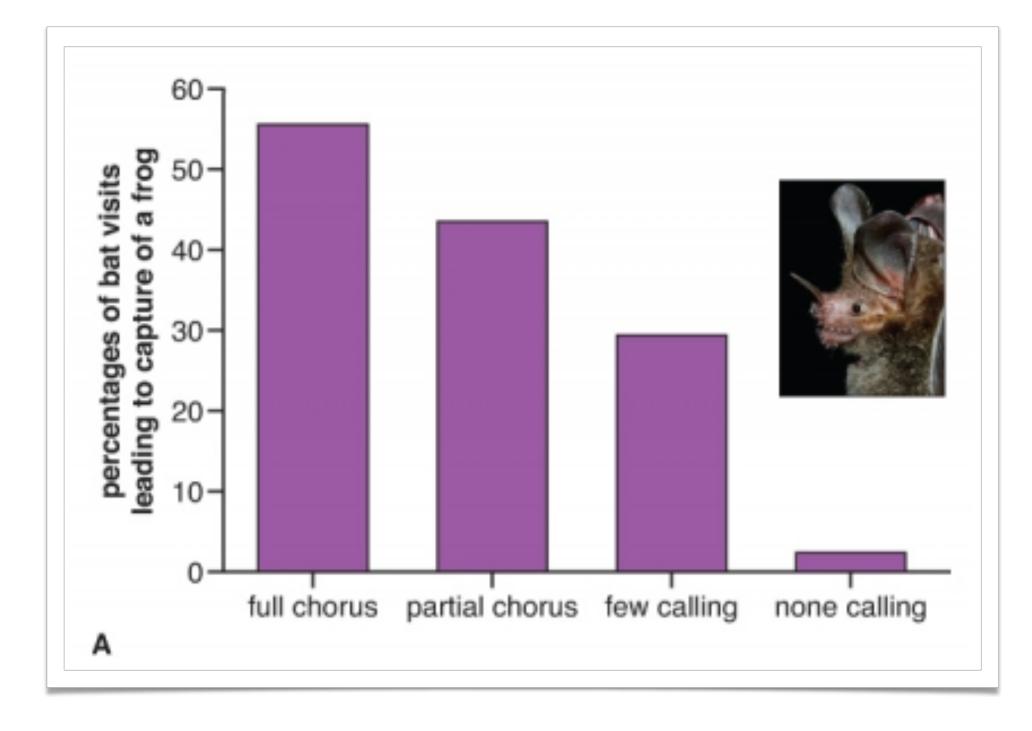
e.g. prepared well for class

These pop quiz questions are designed to reward students who participated,



What's the story with these two animals (which is Correct)?
a) The bats are predators that eat both of them
b) The opossum doesn't eat Tungara frog because is poisonous
c) Both animals live near water and eat bugs and worms
d) The opossum eats small animals and is nocturnal
e) The Tungara frog displays inflatable cheeks to attract females

Figure 18.5 The p The Tungara frog Male. http://upload.w Author: This file is lice Unported license. http://upload.wikimedi pustulosus%29_Callir the Creative Common



because more frogs are there? Nope, there was a speaker with no sounds **a**) Yes, the speaker was nearest to bunch of frogs **b**) Nope, these are not counts of frogs captured C) d)

- Might this data simply be explained by more frogs are caught
 - Yes, a full chorus has more frogs, so it makes sense bats would have a better chance of catching more frogs.

Pop-quiz over

The philander opossum



Figure 18.5

Creative Commons.

The Tungara frog



Figure 18.5

http://www.youtube.com/watch?feature=endscreen&v=5S-RAgudnww&NR=1

Brian Gratwicke, 2012, Creative Commons.



Ryan & Tuttle Methods: opossums observed near a pond that contained breeding male Tungara frogs

Study #1 (purely observational experiment)

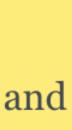
- M: Observed 2 hs/night; night vision scope, saw 39 captures
- F: Typical behavior recorded: describe? **Study #2** (audio playback experiment)
- M: Recordings played if opossum near & facing away from speaker when no frogs vocalizing (Q: speaker placed 2m from pond?!)
- F: 5/5 trials: opossum turned toward speaker, tilted head and rotated ears, approached the speaker, 3/5 trials: pounced on the speaker, continued to circle the speaker if the playback was on. (2 trials: opossum spotted the researchers, and it left)

Integrating Questions

colleagues indicate about the ability of the philander opossum to intercept communication between frogs?



10. Do you think the philander opossum relies more on acoustic signals or visual cues to locate its prey? What do the observations of Ryan and



Prepare for Trifecta (purpose, methods, findings)



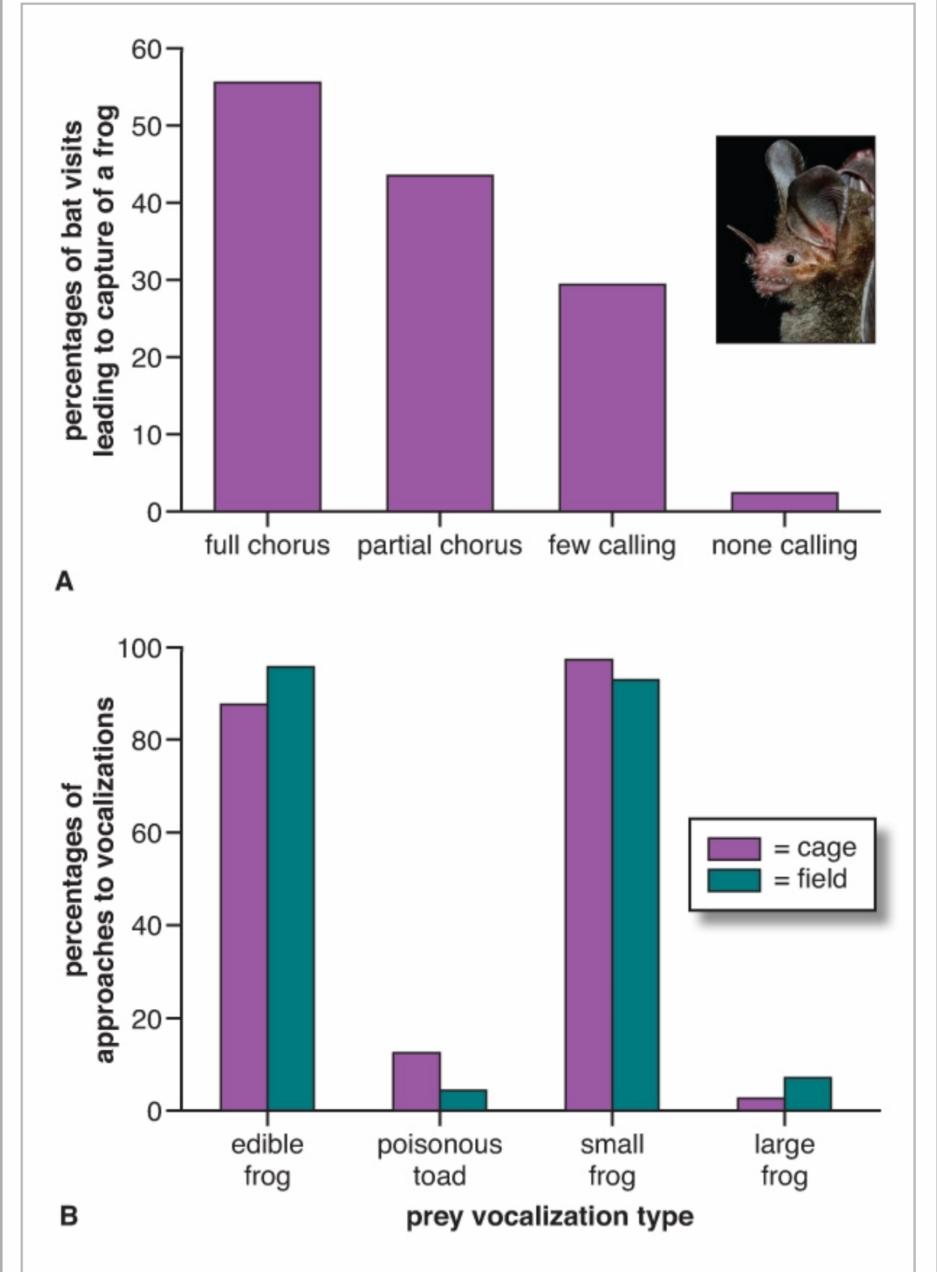
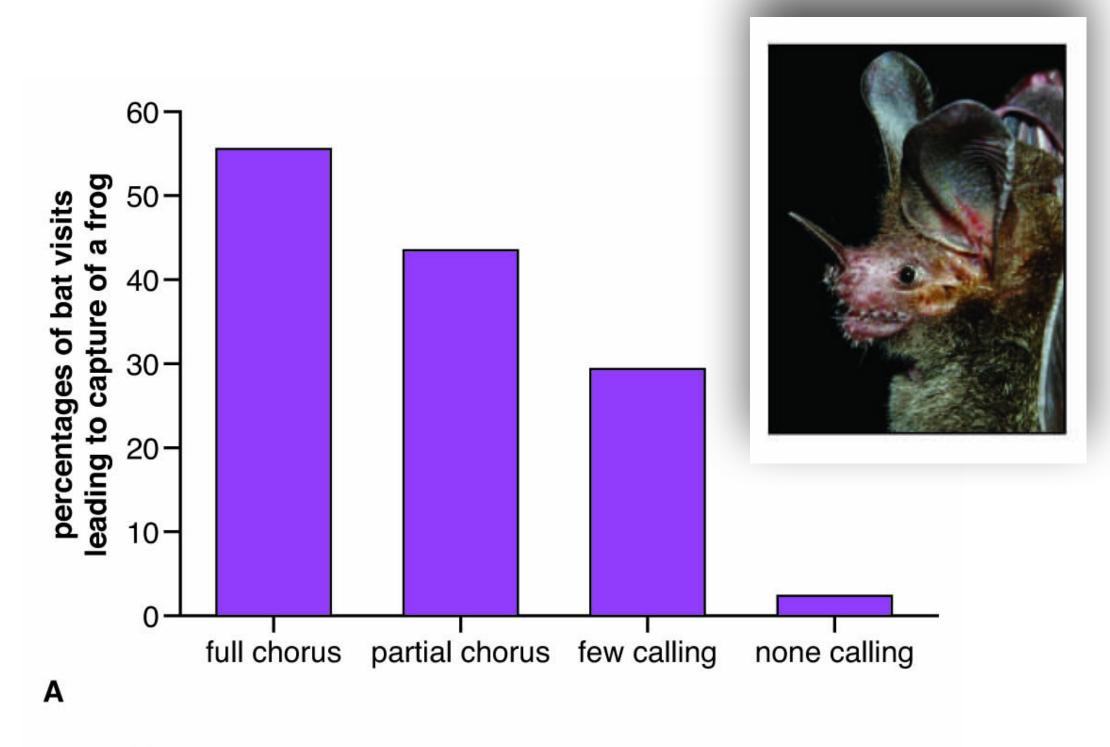


Figure 18.6 Results of experiments on fringe-lipped bats preying on frogs.
A, Percentage of bat visits resulting in a frog capture for four categories of Tungara frog vocalization frequency. Inset, A fringe-lipped bat. B, Responses of frog-eating bats to vocalizations of edible versus poisonous or small versus large frog or toad prey. From Tuttle and Ryan, 1981,



Figure 18.6

From Tuttle and Ryan, 1981, Table 1. Inset: Author: Karin Schneeberger. Creative Commons



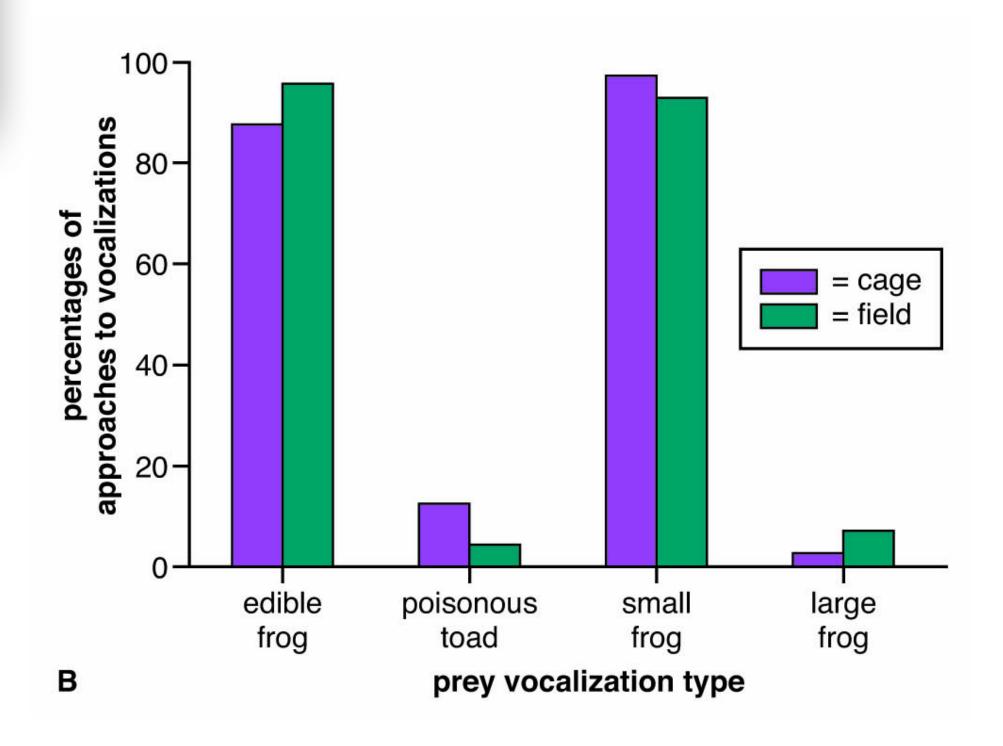
Trifecta? (purpose, methods, findings)



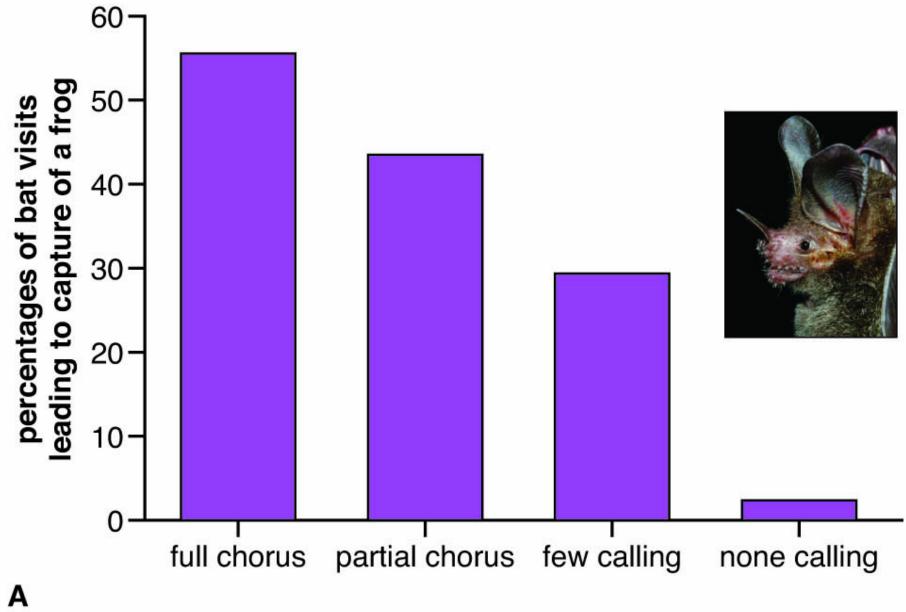
Figure 18.6

From Tuttle and Ryan, 1981, Table 1. Inset: Author: Karin Schneeberger. Creative Commons

Trifecta? (purpose, methods, findings)



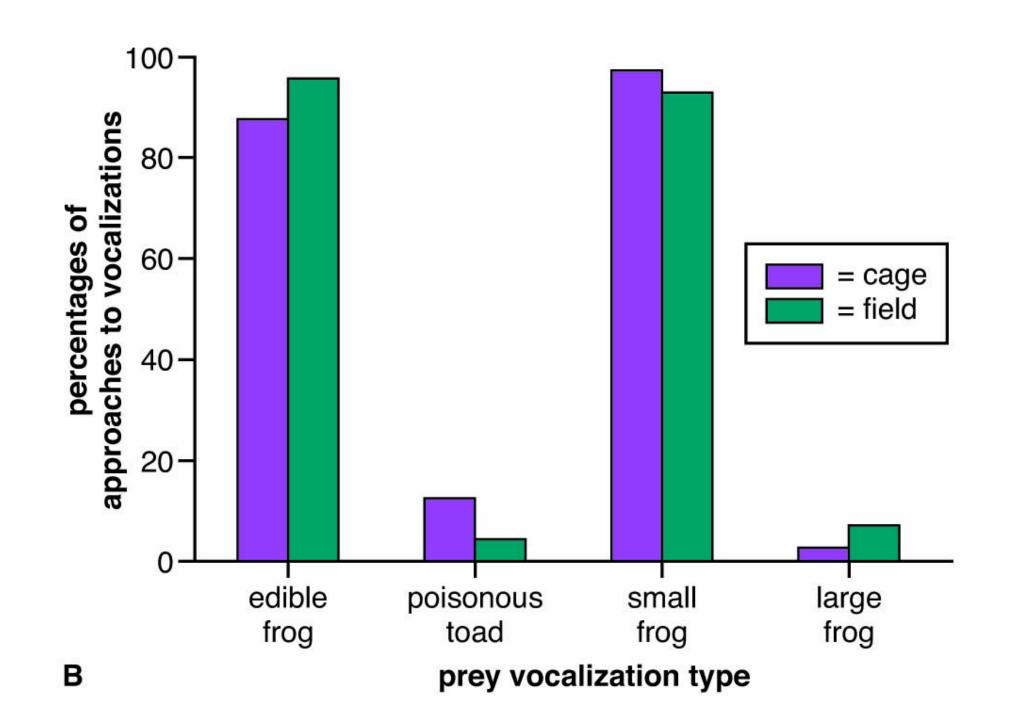
- the bats?
- Which frogs are best protected from predation, despite their vocalizations?
- response for captive bats versus wild bats?



11. What do the success rate data (i.e., the percent of bat visits resulting in a capture) tell you about the hunting methods of

12. Are frog-eating bats able to discriminate among potential prey types? On what basis do you make that conclusion?

13. Why did the researchers perform the experiments both in the cages and in the field? Is there a significant difference in





diminished during hypothyroidism in adult rats. The spontaneous discharge rate of Purkinje neurons is determined to a large extent by the tonic inhibitory noradrenergic afferents originating in the nucleus locus coeruleus (13). Thus, an increased spontaneous discharge in the Purkinje neurons of hypothyroid rats is consistent with the diminished responsiveness of such neurons to norepinephrine. A hypoactive afferent projection from the locus coeruleus is ruled out by several studies on the peripheral and central nervous systems (5, 14, 15) demonstrating an increased amount of norepinephrine discharged at the sympathetic nerve endings during hypothyroidism. Thus, alterations in postsynaptic function could conceivably lead to the electrophysiological changes reported here (6, 15). β -Receptors might be altered in number or in their affinity or functional coupling with adenylate cyclase. In fact, diminished β -receptor binding (with no change in affinity) during hypothyroidism has been demonstrated (3, 15). In peripheral tissues, parallel changes in adenylate cyclase activity have been reported (16). Our electrophysiological studies agree well with previous observations (3, 6, 15, 16). In another study (2), the activity of cyclic AMP phosphodiesterase (E.C. 3.1.4.17) did not change in the cortices of hypothyroid rats. The studies we report here suggest that the subsensitivity occurring

- 9. J. C. Eccles, M. Ito, J. Szentagothai, in *The Cerebellum as a Neuronal Machine* (Springer-Verlag, New York, 1967).
- 10. H. M. Geller and D. J. Woodward, *Electroencephalog. Clin. Neurophysiol.* 33, 430 (1972).
- J. S. Kelly, M. A. Simmonds, D. W. Straughan, in *Methods in Brain Research*, P. B. Bradley, Ed. (Wiley, New York, 1975), p. 333.
- Ed. (Wiley, New York, 1975), p. 333.
 12. For more complete details of this method see R. Freedman and J. Marwaha, *Pharmacol. Exp. Ther.* 212, 390 (1980); J. Marwaha, M. Palmer, B. J. Hoffer, R. Freedman, *ibid.* 215, 606 (1980); J. Marwaha, M. Palmer, B. J. Hoffer, R. Freedman, *ibid.* 215, 606 (1980); J. Marwaha, M. Palmer, B. J. Hoffer, R. Freedman, *ibid.* 215, 606 (1980); J. Marwaha, M. Palmer, B. J. Hoffer, R. Freedman, *ibid.* 215, 606 (1980); J. Marwaha, M. Palmer, B. J. Hoffer, R. Freedman, *ibid.* 215, 606 (1980); J. Marwaha, M. Palmer, B. J. Hoffer, R. Freedman, *ibid.* 215, 606 (1980); J. Marwaha, M. Palmer, B. J. Hoffer, R. Freedman, *ibid.* 215, 606 (1980); J. Marwaha, M. Palmer, B. J. Hoffer, R. Freedman, *ibid.* 215, 606 (1980); J. Marwaha, M. Palmer, B. J. Hoffer, R. Freedman, *ibid.* 215, 606 (1980); J. Marwaha, M. Palmer, B. J. Hoffer, R. Freedman, *ibid.* 215, 606 (1980); J. Marwaha, M. Palmer, B. J. Hoffer, R. Freedman, *ibid.* 215, 606 (1980); J. Marwaha, M. Palmer, B. J. Hoffer, R. Freedman, *ibid.* 215, 606 (1980); J. Marwaha, M. Palmer, B. J. Hoffer, R. Freedman, *ibid.* 215, 606 (1980); J. Marwaha, M. Palmer, B. J. Hoffer, R. Freedman, *ibid.* 215, 606 (1980); J. Marwaha, M. Palmer, B. J. Hoffer, R. Freedman, *ibid.* 215, 606 (1980); J. Marwaha, M. Palmer, B. J. Hoffer, R. Freedman, *ibid.* 215, 606 (1980); J. Marwaha, M. Palmer, B. J. Hoffer, R. Freedman, *ibid.* 215, 606 (1980); J. Marwaha, M. Palmer, B. J. Hoffer, R. Freedman, *ibid.* 215, 606 (1980); J. Marwaha, M. Palmer, B. J. Hoffer, R. Freedman, *ibid.* 215, 606 (1980); J. Marwaha, M. Palmer, B. J. Hoffer, R. Freedman, *ibid.* 215, 606 (1980); J. Marwaha, M. Palmer, B. J. Hoffer, R. Freedman, *ibid.* 215, 606 (1980); J. Marwaha, M. Palmer, B. J. Hoffer, R. Freedman, *ibid.* 215, 606 (1980); J. Marwaha, M. Palmer, B. J. Hoffer, R. Freedman, *ibid.* 215, 606 (1980); J. Marwaha, M. Palmer, B. J. Hoffer, R. Freedman, *ibid.* 215, 606 (1980); J. Marwaha, M. Palmer, B. J. Hoffer, R. Freedman, *ibid.* 215, 606 (1980)
- man, Life Sci. 26, 1509 (1980). 13. B. J. Hoffer, G. R. Siggins, A. P. Oliver, F. E.

Bat Predation and the Evolution of Frog Vocalizations in the Neotropics

Abstract. Bat predation has probably had an important influence on the evolution of frog vocalizations in the Neotropics. The rate at which fringe-lipped bats capture frogs is significantly higher when the frogs are calling. These bats respond to a wide variety of calls from edible frogs, and, when simultaneously presented with a choice, choose the recorded call of a palatable species over that of a poisonous species and the call of a small species over that of one too large to capture. Thus the selective advantages of loud, rapid mating calls in anurans are balanced by an increased risk of predation.

Many animals use conspicuous vocalization to attract mates. The benefits are obvious, but biologists have long suspected that this also leads to increased vulnerability to sound-responsive predators (1, 2). Although such counterselection is believed to influence the evolution of vocal advertisement (3), documentation is rare (2) and is entirely lacking for vertebrates. In this report we show that Bloom, J. Pharmacol. Exp. Ther. 184, 553 (1973).

- 14. S. W. Spaulding and R. H. North, Med. Clin. N. Am. 59, 1123 (1975).
- 15. P. C. Whybrow and A. J. Prange, Arch. Gen. Psychiatry 38, 106 (1981).
- 16. G. S. Levey, C. L. Skelton, S. E. Epstein, J. Clin. Invest. 48, 2244 (1969).
- 17. M. Nakashima and Y. Hagino, Jpn. J. Pharmacol. 22, 227 (1972).
- 18. T. Posternak, E. W. Sutherland, W. F. Henion, Biochim. Biophys. Acta 65, 558 (1962).
- 19. Supported by USPHS grant DA-07043.

10 July 1981

sus) uses acoustic cues to capture calling frogs, and we consider the possible role of call-responsive predators in the evolution of anuran calling and courtship behavior.

On 35 nights from January to June 1980 we visited 14 frog breeding sites on Barro Colorado Island, Panama (4). Trachops cirrhosus was observed hunting on each night. Seven of the nights were

| Species | In cage | | In field | | |
|--|------------------------------------|---------------------|--|--|--|
| | Bats tested | Responses | Sites visited | Responses | |
| Hyla boulengeri | 5 | 35 | 6 | 66 | |
| Bufo typhonius | 5 | 5 | 6 | 3 | |
| x^2 | 42.81 (10), P < .005 | | 70.63 (12), $P < .005$ | | |
| Physalaemus pustulosus | 5 | 36 | 3 | 26 | |
| Leptodactylus pentadactylus | 5 | 1 | 3 | 2 | |
| χ^2 | 47.15 (10), <i>H</i> | 47.15(10), P < .005 | | 31.52 (6), $P < .005$ | |
| • | | | a mining at the law and the standard and shared the second fill the standard the second is a supply supply a | ی دوستهای و میکند. این | |
| Table 2. Responses of T. cirrho | | | ent calls of two an | nuran species | |
| | | | | nuran species esponses | |
| played at different repetition rates Species "calling" | | • | | | |
| played at different repetition rates Species "calling" | tes and volumes | • | | | |
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Table 1. Responses of T. cirrhosus to playbacks of the advertisement calls of four anuran

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| treatment | mean number of trials | standard error | sample size |
|---|--------------------------|-------------------|----------------|
| inexperienced bat with experienced bat | 5.3 | 1.7 | 10 |
| two inexperienced bats | 96.8 | 3.2 | 5 |
| one inexperienced bat | 96.2 | 3.8 | 5 |

Table 18.2. Number of trials needed for fringe-lipped bats to learn to associate the cane toad vocalization with palatable prey. From Page and Ryan, 2006, Figure 2.

Prepare for Trifecta (purpose, methods, findings)

What was the next experiment?

Trifecta

| treatment | mean number of trials | standard error | sample size |
|---|--------------------------|-------------------|----------------|
| inexperienced bat with experienced bat | 5.3 | 1.7 | 10 |
| two inexperienced bats | 96.8 | 3.2 | 5 |
| one inexperienced bat | 96.2 | 3.8 | 5 |

(purpose, methods, findings)

| treatment | mean number of trials | standard error | sample size |
|---|--------------------------|-------------------|----------------|
| inexperienced bat with experienced bat | 5.3 | 1.7 | 10 |
| two inexperienced bats | 96.8 | 3.2 | 5 |
| one inexperienced bat | 96.2 | 3.8 | 5 |

14. What can you conclude from the data in Table 18.2? Do the data support what you expected?15. Identify the controls that Rachel Page and Mike Ryan used in their experiment and what the controls were for.

Social Transmission of Novel Foraging Behavior in Bats: Frog Calls and Their Referents

Rachel A. Page^{1,2,*} and Michael J. Ryan^{1,2} ¹ Section of Integrative Biology University Station C0930 University of Texas at Austin Austin, Texas 78712 ² Smithsonian Tropical Research Institute Apartado 2072 Balboa Republic of Panama

Summary

The fringe-lipped bat, *Trachops cirrhosus*, uses preyemitted acoustic cues (frog calls) to assess prey palatlarge for a *T. cirrhosus* to eat, so on two accounts it should be an unsuitable prey item. The criterion for ability [1]. Previous experiments show that wild *T. cirrhosus* brought into the laboratory are flexible in their task acquisition was flying to and landing on a speaker ability to reverse the associations they form between broadcasting toad calls in three consecutive trials. prey cues and prey quality [2]. Here we asked how We first conducted baseline tests with all bats to dethis flexibility can be achieved in nature. We quantified termine initial responses to *B. marinus* calls. None of the rate at which bats learned to associate the calls of the bats showed any initial response to *B. marinus* calls. We then tested for social learning by allowing an inexpea poisonous toad species with palatable prey by placing bats in three groups: (a) social learning, in which rienced bat to observe the foraging behavior of an expea bat inexperienced with the novel association was alrienced bat (tutor) that had already acquired the novel association. The first tutor learned to associate toad lowed to observe an experienced bat; (b) social facilitation, in which two inexperienced bats were precalls with a palatable food reward via a fading-condisented with the experimental task together; and (c) tioning technique (for methods see [2]) Subsequent tu-

Report

prey palatability [2]. Using a fading-conditioning paradigm [6], we were able to rapidly reverse the bats' assessment of palatable and poisonous prey.

Here we ask whether this flexibility is part of the bats' natural foraging repertoire and to what degree novel associations between prey cue and prey quality can be culturally transmitted. To address these questions, we quantified the rate of acquisition of a novel foraging behavior in three learning groups: (a) a social-learning group, (b) a social-facilitation group, and (c) a trial-and-error group. The target foraging behavior was the bats' ability to learn to associate the calls of the sympatric cane toad, *Bufo marinus*, with a palatable food reward. *B. marinus* is both highly poisonous and far too large for a *T. cirrhosus* to eat, so on two accounts it should be an unsuitable prey item. The criterion for task acquisition was flying to and landing on a speaker broadcasting toad calls in three consecutive trials.

strate that mothers and their daughters shared foraging grounds, sometimes for years [27, 28]. Thus, the vertical transfer of foraging-site location from mother to pup could be playing a large role in the foraging dynamics of these bat communities. Although the learning we document in our study is likely entirely opportunistic (the result of one bat eavesdropping on the successful foraging behavior of another), the study of social learning in highly related groups, and especially in mother-pup pairs, should prove an interesting area for further research.

Our study is not designed to distinguish among the mechanisms of social learning [29–32]; however, it is likely that these bats are learning by either stimulus enhancement or observational conditioning. In stimulus enhancement the activity of the tutor draws the observer's attention to the test stimulus [31, 33]—in our experiment, to the toad calls. The observer then forms an association between the stimulus and the reward via individual, trial-and-error learning. Because we altered the speaker location for each trial, we can rule out the possibility that the bats are learning to associate a food reward with a particular spatial location (local enhancement).

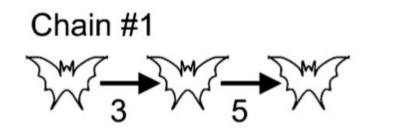
In observational conditioning, a type of higher-order conditioning, the observer associates the stimulus with the outcome experienced by the tutor and thus responds more readily to the stimulus itself [31, 34]. In our social-learning treatment, the test bat did not initially attend to the toad calls or to the flight of the tutor bat. In the initial trials, the test bat typically would commence responding with ear motions and head orientation only

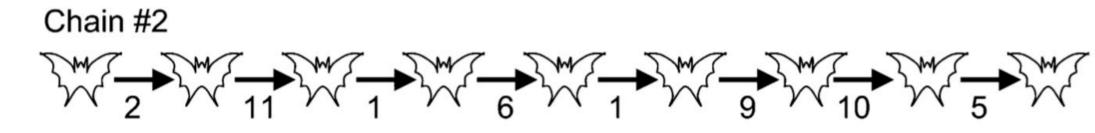
Experimental Procedures

Experiments were conducted at the Smithsonian Tropical Research Institute field station on Barro Colorado Island (BCI), Panama, from February to June 2004 and 2005. We captured the bats in mist nets and tested them in a 4.5 m × 4.5 m × 2.5 m outdoor flight cage. We illuminated the flight cage with a 25 watt red light bulb to facilitate our observations of the bats. This light level was within the range of illuminations in which the bats forage. We used a Sony NightShot DCR-TRV340 camera equipped with a Sony HVL-IRH2 infrared light to record all initial and final tests, all social learning trials, and a subset of the social-facilitation and trial-and-error learning trials. Each bat was marked with a passive integrated transponder (PIT tag) and released at its site of capture after testing. All experiments were licensed by the Smithsonian Tropical Research Institute and the University of Texas at Austin (IACUC #04113002).

Stimulus Presentation

We broadcast calls of B. marinus from a Dell Inspiron 8100 computer, a SA-150 Realistic amplifier, and 40-1040 Radio Shack speakers. To approximate the natural call intensity of *B. marinus* in the wild, we broadcast the calls at an amplitude of 75 dB SPL (re. 20 μ P) measured at a distance of 1 m from the speaker. Most of the energy in *B. marinus* calls falls between 548 and 708 Hz; the frequency response of these speakers is flat for these frequencies. To ensure that the bats responded to the acoustic stimulus broadcast and not to the speaker itself, we concealed one to five speakers beneath a $1.5 \text{ m} \times 1.5 \text{ m}$ screen covered with leaf litter and randomly repositioned the speakers between trials. To ensure that the bats were responding to the toad calls per se and not to other noises associated with the speaker, in a subset of the trials we turned on one of the control speakers and broadcast a sound file of silence. The bats never approached control speakers. Toad calls were broadcast for 60 s or until the test bat landed on the speaker, whichever came first. Trials were conducted in approximately 10 to 15 min intervale with a maximum of 20 trials nor night





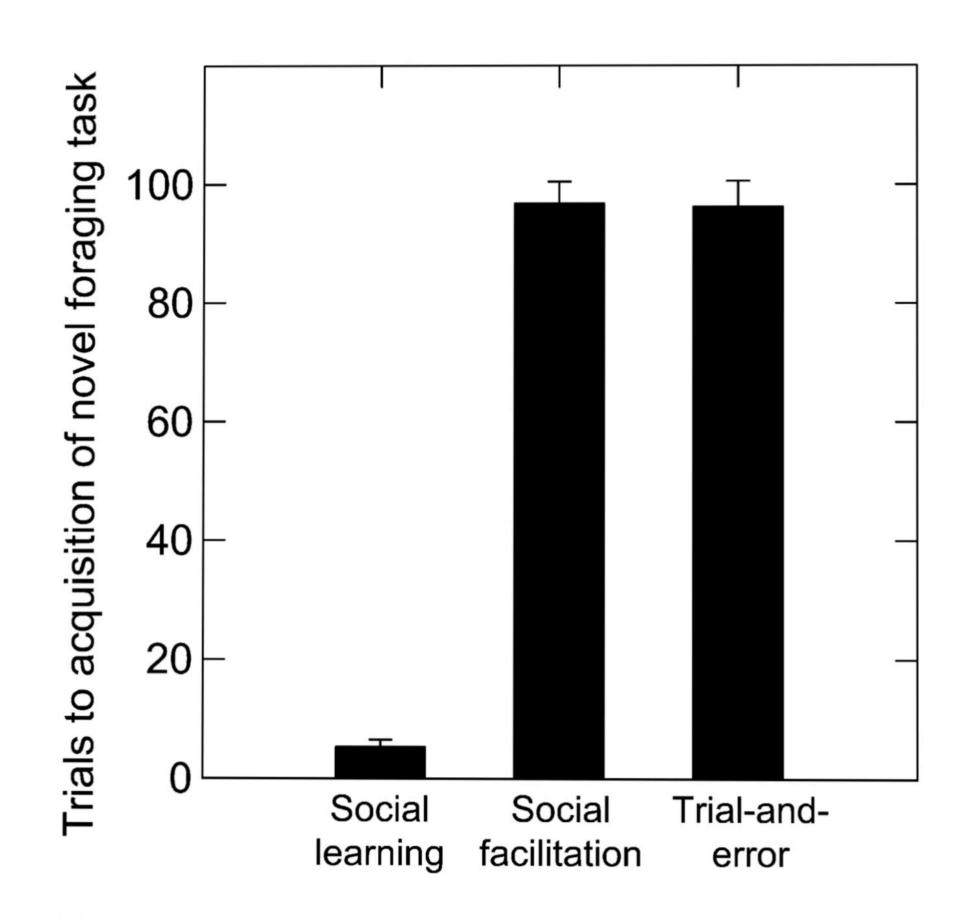


Figure 2. Mean Rates of Acquisition, ± SEM, of the Novel Foraging Task for the Three Learning Groups

Figure 1. Chains of Transfer in the Social Learning Group

Arrows indicate the transmission of the novel foraging behavior; the numbers underneath the arrows indicate the number of trials required to reach acquisition criterion for each bat. After testing, the current test bat became the tutor for the next test bat.

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